PRIMARY RESEARCH PAPER

Differences in leaf nitrogen content, photosynthesis, and resource-use efficiency between Eichhornia crassipes and a native plant *Monochoria vaginalis* in response to altered sediment nutrient levels

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Abstract Eichhornia crassipes is one of the world's most prevalent invasive aquatic plants, causing significant ecological and socio-economic impacts in introduced areas. In this study, we compared the leaf resource capture- and use-related traits of E. crassipes with its confamilial native aquatic plant Monochoria vaginalis at three nutrient levels. Our results showed that leaf nitrogen content based on mass, leaf nitrogen content based on area, N:C, photosynthetic rate, specific leaf area, and leaf construction cost of E. crassipes increased significantly with increasing levels of nutrition, the mean values of these traits increased 0.55, 0.35, 0.51, 0.43, 0.21, and 0.07 times from low nutrient level to high, respectively. These traits (except for the leaf construction cost) in M. vaginalis remained unchanged. At low nutrient level, M. vaginalis had a higher leaf nitrogen content, N:C, photosynthetic rate, specific leaf area, and water-use efficiency than E. crassipes. At high nutrient level, E. crassipes had a higher photosynthetic rate and photosynthetic nitrogen-

Introduction

crassipes.

Invasive species degrade human health (Juliano & Lounibos, 2005), cause huge economic losses (Pimentel et al., 2005), reduce biodiversity, and alter species composition, structure, processes, and function of the invaded ecosystems (Vitousek, 1990; D'Antonio & Kark, 2002; Svilà et al., 2011). Unfortunately, due to an increase in international commerce, more alien species have been introduced into new habitats (Cohen & Carlton, 1998; Hulme, 2009). In addition, human activities and global changes (such as changes in land-use pattern, nitrogen deposition, and increased atmospheric CO₂) have facilitated such invasions and enhanced their adverse effects (Dukes & Mooney, 1999). Therefore, to improve our ability to control existing invasive species and forecast

use efficiency than M. vaginalis, suggesting that the

invasiveness of E. crassipes was dependent on the

availability of resources in environment. In addition, our results supported the fluctuating resources hypoth-

esis, indicating that an increased level of nutrients in the

environment will increase the invasiveness of E.

Keywords Biological invasion · Water hyacinth ·

Photosynthesis · Resource-use efficiency ·

Fluctuating resources hypothesis

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potentially invasive species, it is necessary to determine the mechanisms of invasion and identify the traits that increase the invasiveness of a species.

Resource-use efficiency affects the growth, reproduction, and competitiveness of plants. Vitousek (1986) proposed that, if the life history traits and susceptibility to herbivore pressure are similar between an invasive and a native species, the successful invasive species must either use limited resources more efficiently than the native species or use them at times when they are unavailable to the native species. The traits commonly linked to invasiveness in plants include higher relative growth rate (Pattison et al., 1998; Burns, 2004; Garcia-Serrano et al., 2005; Grotkopp & Rejmánek, 2007; Xie et al., 2010), higher photosynthetic rate (Pattison et al., 1998; Baruch & Goldstein, 1999; Durand & Goldstein, 2001; McDowell, 2002; Funk & Vitousek, 2007), larger specific leaf area (Grotkopp & Rejmánek, 2007; Feng et al., 2008; Osunkoya et al., 2010), lower leaf construction cost (Baruch & Goldstein, 1999; Feng et al., 2007; Osunkoya et al., 2010), and higher photosynthetic nitrogen-use efficiency (Niinemets et al., 2003; Feng et al., 2007; Feng & Fu, 2008; Matzek, 2011). All of these traits are associated with better resource-use strategy, and some traits expressed together to facilitate invasion.

Resource availability can also influence the resourceuse strategy of plants. In a resource-rich environment, plants may have a higher relative growth rate, photosynthetic rate and leaf nutrient concentration, while in a resource-poor environment, plants may have a lower relative growth rate, photosynthetic rate, and leaf nutrient concentration (Chapin, 1980; Bloom et al., 1985). Many studies have found that it is rare for invasive plants to consistently outperform native plants; only in resource-rich environments do invasive plants display advantageous traits such as a higher relative growth rate, photosynthetic rate, and specific leaf area (Daehler, 2003; Burns, 2004). Davis et al. (2000) proposed the fluctuating resources hypothesis and concluded that a plant community becomes more susceptible to invasion when there is either an increase in the amount of unused resources or a decreased uptake of the resources by the native plant community. Accordingly, invasive plants have higher resource-use efficiency than native plants only in resource-rich habitats. In contrast, comparing 19 pairs of phylogenetically related invasive and native species, Funk &

Vitousek (2007) found that the invasive plants had a higher instantaneous photosynthetic capacity and photosynthetic nitrogen-use efficiency than the native plants in three habitats in Hawaii in which the light, water, or nutrient availability was limiting to plant growth. Matzek (2011) also found that five invasive pines had a higher relative growth rate, photosynthetic capacity, and photosynthetic nitrogen-use efficiency than five non-invasive pines in a nutrient-limiting environment. Owing to the inconsistent results for the resource-use efficiency of invasive and native plants under different resource conditions, Funk & Vitousek (2007) suggested that more data are needed regarding how the patterns of resource acquisition and usage vary in response to fluctuating resources between invasive and native plants. Some researchers have compared many resource capture- and use-related traits of invasive and native plants under single conditions (Baruch & Goldstein, 1999; Feng et al., 2007; Funk & Vitousek, 2007; Feng & Fu, 2008; Matzek, 2011), but few studies investigated the different patterns of these traits in response to changes in the resource availability. In addition, the plants being studied were all terrestrial plants.

Eichhornia crassipes (Mart.) Solms (Water hyacinth) is one of the world's most prevalent invasive aquatic plants in the monocotyledonous family Pontederiaceae, having significant ecological and socioeconomic impacts in areas of introduction (Villamagna & Murphy, 2010). Many studies have been conducted on E. crassipes, mostly focusing on the effect, control, and utilization of the plant, instead of the invasion mechanism. Although some researchers have studied the invasion mechanism of E. crassipes (Xie & Yu, 2003; Xie et al., 2004), few studies have compared its traits with phylogentically related plants. Indeed, comparisons with a closely related species are difficult because E. crassipes is the only floating species in the family Pontederiacea. Other studies (Parker & Hay, 2005; Xiong et al., 2008) have compared the native herbivore preference for E. crassipes with the confamilial aquatic emergent plants Pontederia cordata L. and Monochoria vaginalis (Burm.f.) Presl, native in North America and Asia, respectively. Although E. crassipes is normally a free-floating plant, in some habitats, such as flooded sites, swamps and at the edges of pools, ditches, lakes, and canals, it can root and grow as an emergent plant (Njambuya & Triest, 2010). During seedling stage, E. crassipes must root in muddy



banks of waters or in rice field. Because *E. crassipes* has similar leaf structure with *M. vaginalis*, we compared the leaf traits of these two species at three nutrient levels to address the following questions: (i) Does *E. crassipes* differ from *M. vaginalis* in resource capture- and userelated traits? (ii) Does *E. crassipes* display different morphological and physiological patterns compared with *M. vaginalis* in response to changes in nutrient levels? (iii) Does *E. crassipes* achieve a higher photosynthesis and resource-use efficiency than *M. vaginalis* only at high nutrient level?

Materials and methods

Study plants

Originally from South America, E. crassipes has invaded more than 62 countries in the tropical, subtropical, and temperate zones of Africa, Asia, North America, and Oceania (Howard & Harley, 1998). E. crassipes reproduces both sexually and asexually (data not published) but is spread primarily by vegetative propagation in China. Under the appropriate environmental conditions, E. crassipes can reproduce a new ramet in 5 days by cloning; a single plant can produce 140 million daughter plants (28,000 tons of fresh weight) every year, sufficient to cover 140 ha (Ogutu-Ohwayo et al., 1997). Therefore, E. crassipes has been considered one of the fastest growing plants on the earth (Abbsi & Nipaney, 1986). In invaded areas, E. crassipes can change the hydrological environment, destroy the native animal and plant communities, and reduce the species diversity, thus altering the ecosystem structure and function by disrupting the food chain and nutrient cycling (Villamagna & Murphy, 2010). Accordingly, E. crassipes has been listed as one of the 100 most dangerous invasive species by IUCN (Lowe et al., 2000).

The native annual aquatic emergent plant *M. vaginalis* also belongs to the Pontederiaceae family and is distributed in Japan, Malaysia, Philippines, India, Nepal, Bhutan, and the northern and southern provinces of China. *M. vaginalis* is often gregarious and is typically found in flooded sites, swamps, rice fields, and edges of pools, ditches, and canals. *M. vaginalis* reproduces by sexual propagation, flowering in August and September, and fruiting in September and October in China.

Experimental design

This experiment was conducted in The National Field Station of Freshwater Ecosystem of Liangzi Lake, Hubei Province, China (30°50′-30°180′N, 114°210′-114°390′E). A total of 40 clonal ramets of E. crassipes and 40 seedlings of M. vaginalis were collected in a field population of Liangzi Lake at the end of July. The plants were cultivated in eight circular basins of 36 cm diameter with a sand sediment and 5 cm water depth. One week later, 18 plants of each species with similar weight and height (the mean fresh weight of E. crassipes was 8.44 ± 0.36 g, and M. vaginalis was 6.61 ± 0.56 g) were transferred to 36 rectangular basins (36 cm length \times 30 cm width \times 15 cm height) filled with approximately 8 cm of sand (from the banks of Liangzi Lake, N:P = 0.13:0.014mg g⁻¹) and 7 cm depth of water (from Liangzi Lake, $N:P = 0.71:0.04 \text{ mg L}^{-1}$). All plants were rooted in sand, and grown under three nutrient levels, with six replicas each. To the high and medium nutrient treatments, 10 and 5 g, respectively, of slow-release fertilizer (containing 16 g N and 8 g P per 100 g) were mixed in sand; no fertilizer was added to the low nutrient treatment. The nitrogen concentration of the three nutrient levels were 0.06, 0.14, and 0.22 mg g^{-1} , similar to the sand, soil, and plant litter sediments of Liangzi Lake, respectively. To facilitate the management of the experiment, six plants in the same treatment were arranged as a group, and six groups were randomly placed on an outdoor cement platform (10 m length × 8 m width). All plants were exposed to the same intensity and duration of light and were watered daily to maintain a constant water level until they were harvested.

Measurements

At 40 days after planting, all plants (except for *E. crassipes* at the low nutrient level) started inflorescence stage, and their leaf traits were measured (Table 1). The net photosynthetic rate (Pn) and stomatal conductance (Gs) was determined on the youngest fully expanded leaves using a Li-6400 Portable Photosynthesis System (Li-Cor, USA) under a photosynthetic photon flux density (PPFD) of natural light >1,700 μ mol m⁻² s⁻¹, the air temperature was moderate (25–35°C) and the relative humidity ranged between 60 and 70% at 11:00 a.m. to 2:00 p.m. The



Table 1 Definitions and abbreviations of measured traits

Traits	Description	Units
$N_{ m mass}$	Leaf nitrogen content based on mass	%
$N_{\rm area}$	Leaf nitrogen content based on area	$\mathrm{g}~\mathrm{m}^{-2}$
N:C	Ratio of leaf nitrogen and carbon content on mass	
P_{max}	Maximum photosynthetic rate at saturating light	$\mu mol\ m^{-2}\ s^{-1}$
SLA	Leaf area per mass	$\mathrm{cm}^2~\mathrm{g}^{-1}$
LCC	Leaf construction cost	$\mathrm{g}~\mathrm{cm}^{-2}$
PUNE	Photosynthetic nitrogen-use efficiency	$\mu mol~g^{-1}~s^{-1}$
WUE	Water-use efficiency	$\underset{s^{-1}}{\mu mol}\ mmol^{-1}$

leaves used for the photosynthesis measurements were detached, and leaf area was measured using a Li-3100 Area Meter (Li-Cor, USA) to calculate the light-saturated photosynthetic rate per unit area leaf (P_{max}) . From each leaf, 3–5 leaf squares of 1 cm² were punched and immediately dried at 70°C for more than 48 h. One leaf square was used to determine the nitrogen content (N_{mass}) and carbon content (C_{mass}) using an element analyzer Euro EA3000 (Euro Vector, Italy). The remaining leaf squares were used to calculate the specific leaf area (SLA). N_{area} was calculated as N_{mass}/SLA. N:C was calculated as $N_{\rm mass}/C_{\rm mass}$. Photosynthetic nitrogen-use efficiency (PNUE) was calculated as $P_{\text{max}}/N_{\text{area}}$. Water-use efficiency (WUE) was calculated as the ratio of P_{max} to Gs. Leaf construction cost (LCC) was calculated according to McDowell (2002).

Statistical analyses

All traits were analyzed using factorial ANOVA, then Duncan tests were used to compare levels within factors for significance (P < 0.05). All analyses were performed using SPSS 13.0 (SPSS Inc., Chicago, IL, USA).

Results

Differences of traits in species

 $N_{\rm mass}$, $N_{\rm area}$, N:C, and WUE differed significantly between species and across sediment nutrient levels. The significant interactions between nutrients and

species indicated that differences in $N_{\rm mass}$, N:C, and WUE between the two species were significantly influenced by sediment nutrients (Table 2). $N_{\rm mass}$ and N:C of *E. crassipes* were lower than those of *M. vaginalis* at each nutrient level (Fig. 1a, c). At the low nutrient level, *E. crassipes* displayed a lower $N_{\rm area}$ and WUE than *M. vaginalis*, but there were no significant differences compared with *M. vaginalis* at the medium or high nutrient levels (Fig. 1b, g).

Table 2 F and P values of the leaf traits for the two species and three nutrient levels (a factorial ANOVA)

Source	df	F	P
$N_{ m mass}$			
Species	1.6	90.465	< 0.001
Nutrient	2.6	9.673	0.001
Species × nutrient	2.6	6.481	0.005
$N_{\rm area}$			
Species	1.6	6.402	0.017
Nutrient	2.6	10.538	< 0.001
Species × nutrient	2.6	1.277	0.294 ns
N:C			
Species	1.6	79.667	< 0.001
Nutrient	2.6	11.664	< 0.001
Species × nutrient	2.6	4.524	0.019
$P_{\rm max}$			
Species	1.6	0.665	0.421 ns
Nutrient	2.6	25.833	< 0.001
Species × nutrient	2.6	10.836	< 0.001
SLA			
Species	1.6	72.021	< 0.001
Nutrient	2.6	1.037	0.367 ns
Species × nutrient	2.6	6.744	0.004
LCC			
Species	1.6	82.19	< 0.001
Nutrient	2.6	0.39	0.681 ns
Species × nutrient	2.6	29.828	< 0.001
WUE			
Species	1.6	6.628	0.017
Nutrient	2.6	26.575	< 0.001
Species × nutrient	2.6	3.848	0.036
PNUE			
Species	1.6	9.979	0.004
Nutrient	2.6	0.32	0.728 ns
Species × nutrient	2.6	1.001	0.379 ns

Boldface denotes significance, ns denotes no significance

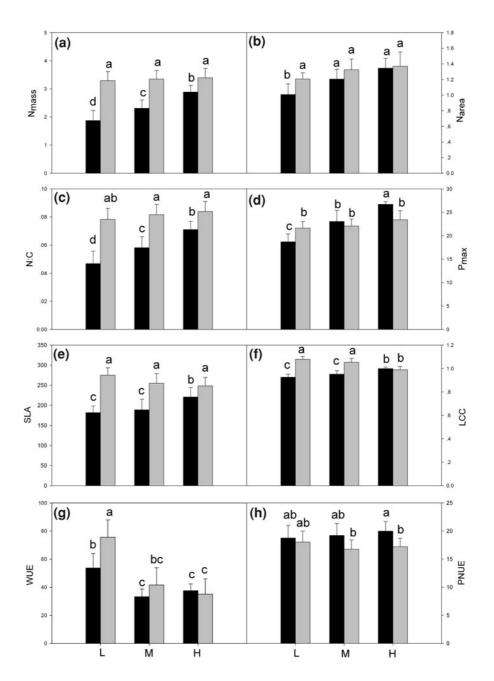


There were no significant differences in $P_{\rm max}$ between the two species. However, among different sediment nutrient levels, $P_{\rm max}$ differed significantly. The significant interactions between nutrients and species suggest that sediment nutrients contribute to the differences of $P_{\rm max}$ between the two species (Table 2). At the low nutrient level, M. vaginalis had a higher $P_{\rm max}$ than E. crassipes, but these differences

become not significant or even reversed at medium and high nutrient levels (Fig. 1d).

SLA, LCC, and PNUE all differed significantly between species, although they remained similar among different sediment nutrient levels (Table 2). SLA of *E. crassipes* was lower than *M. vaginalis* at each nutrient level (Fig. 1e). LCC of *E. crassipes* was lower than *M. vaginalis* at low and medium

Fig. 1 The mean values of *E. crassipes* (*black bars*) and *M. vaginalis* (*gray bars*) in leaf traits at three nutrient levels (*L* low nutrient level, *M* medium nutrient level, *H* high nutrient level)





nutrient levels, but was comparable to *M. vaginalis* at high nutrient level (Fig. 1f). Although PNUE differed significantly between the two species, it is much higher only at high nutrient level in *E. crassipes* than in *M. vaginalis* (Fig. 1h).

Trait response patterns to nutrient

As shown in Fig. 1, most traits of the two species differ in response to changes in nutrient levels. $N_{\rm mass}$, $N_{\rm area}$, N:C, $P_{\rm max}$, and SLA of *E. crassipes* increased significantly when the nutrient level increased, whereas those of *M. vaginalis* remained constant at all three nutrient levels. LCC of *E. crassipes* also increased significantly when the nutrient level increased. In contrast, this trait of *M. vaginalis* decreased significantly with increases in the nutrients. WUE of both species decreased significantly when the nutrient level increased, PNUE of both species remained constant at all three nutrient levels.

Discussion

In our study, the resource capture- and use-related traits were markedly different between E. crassipes and M. vaginalis. M. vaginalis had a higher N_{mass} than E. crassipes at all three nutrient levels. N_{area} of M. vaginalis was also higher than E. crassipes at low nutrient level. Previous studies have found that compared with native plants, the invasive plants generally have higher N_{mass} (Baruch & Goldstein, 1999; Leishman et al., 2007) and lower or similar N_{area} (Baruch & Goldstein, 1999; Feng et al., 2007; Leishman et al., 2007; Feng & Fu, 2008). Exceptions, in which N_{mass} of invasive plants was lower or equal to native plants, did exist (Baruch & Goldstein, 1999; Durand & Goldstein, 2001). Because proteins participating in the Calvin cycle and in the thylakoids represent the majority of the leaf nitrogen content, photosynthesis of a leaf correlate positively to these proteins, hence the leaf nitrogen content (Evans, 1989). However, a lower leaf nitrogen content can partially contribute to a high PNUE in invasive plants, owing to an inverse correlation between nitrogen-use efficiency and foliar nitrogen content (Niinemets et al., 2003). Therefore, consistent with this finding, we discovered that E. crassipes with a lower leaf nitrogen content achieved a higher PNUE than *M. vaginalis*.

SLA and LCC are two other important traits representing relative growth rate and resource-use efficiency in plants (Shipley, 2006). Plants with a high SLA and low LCC can produce larger assimilatory surfaces for a given amount of fixed carbon (Baruch & Goldstein, 1999). Furthermore, plants with a high SLA often allocate a higher fraction of the leaf nitrogen to photosynthesis, resulting in a higher PNUE than plants with a low SLA (Poorter & Evans, 1998; Feng et al., 2008). Many researchers found that, compared with native plants, invasive plants often have a higher SLA and lower LCC (Baruch & Goldstein, 1999; Grotkopp et al., 2002; Grotkopp & Rejmánek, 2007; Feng et al., 2007; Feng & Fu, 2008; Feng et al., 2008). However, in our experiment, we found that E. crassipes had a lower LCC than M. vaginalis only at the low and medium nutrient levels. E. crassipes also had a lower SLA than M. vaginalis at all three nutrient levels. McDowell (2002) suggested that a lower SLA in invasive species may increase the WUE, which means these plants had thicker, denser leaves, increasing the distance through which water must diffuse thereby leading to water conservation. Since the water was not a limiting factor for these aquatic plants in our study, we suggested that a higher SLA was not always a necessary trait for invasive species. This is consistent with some studies in which SLA of invasive plants was found to be lower than native plants (McDowell, 2002; Feng, 2008).

Many invasive plants were found to have higher P_{max} values than their phylogenetically related native species (Pattison et al., 1998; Funk & Vitousek, 2007; Leishman et al., 2007; Feng & Fu, 2008). Photosynthetic rate is related to various attributes of plant success, including growth, competitiveness, propagation, and resistance to herbivores or pathogens (McAllister et al., 1998). Therefore, P_{max} is an important leaf trait of invasive plants. In our study, E. crassipes exhibited a higher P_{max} than M. vaginalis only at the high nutrient level, suggesting that it can obtain more energy and achieve higher growth rate to compete with native plants when resources are abundant. Although PNUE of E. crassipes was higher compared with M. vaginalis, the difference was significant only at the high nutrient level. In introduced areas, due to release from natural enemies, invasive plants will allocate more nitrogen to their



photosynthetic machinery by reducing the nitrogen of cell walls to achieve a higher photosynthetic rate (Feng et al., 2009). In the field, nitrogen is a limiting resource, plants with high PNUE can achieve a higher photosynthetic rate when their leaf nitrogen content is low, it is important to the growth and propagation, therefore, many invasive plants were found to have higher PNUE than their phylogenetically related native and non-invasive species (Niinemets et al., 2003; Feng et al., 2007; Funk & Vitousek, 2007; Matzek, 2011).

There are many different morphological and physiological response patterns to changes in nutrition between E. crassipes and M. vaginalis. Generally, plants adapted to resource-poor habitats demonstrate a low growth rate, tissue nutrient content, instantaneous photosynthetic rate, and instantaneous PNUE, but exhibit a long leaf lifespan and greater leaf thickness, allowing them to have a longer time to maximize their carbon assimilation per unit resource invested in leaf construction (Chapin, 1980, Bloom et al., 1985). In our study, E. crassipes had a decreased photosynthetic rate, leaf nitrogen content, and SLA with decreasing nutrient levels. In contrast, M. vaginalis maintained steady values for P_{max} , N_{mass} , N_{area} , N:C, and SLA at all the nutrient levels. In the Liangzi Lake, E. crassipes reproduces mainly by clonal propagation, though a few seedlings also were found. Conversely, M. vaginalis reproduces by sexual propagation. The carbon and nutrient cost for producing reproductive structures (flowers, fruit, and seed) is generally high because of the high concentrations of nitrogen, phosphorus, and lipids in these organs (Bloom et al., 1985). Therefore, compared with asexual reproduction, sexual reproduction requires more resources from the environment. In a resource-poor environment, plants will increase asexual reproduction and decrease sexual reproduction. For the annual sexually reproducing aquatic plant M. vaginalis, a high P_{max} is necessary to support the high energy cost of reproductive structures. Thus, M. vaginalis must maintain a high leaf nitrogen concentration to sustain a high photosynthetic rate and a high SLA and LCC to increase its light-capture ability in low nutrient level.

The fact that most traits of *E. crassipes* increased with increasing nutrition suggest that it favors nutrient-rich habitats. Many studies have found that some extremely invasive aquatic weeds tend to increase their biomass, clonal propagation, and propagule

rooting efficiency in high nutrient environments (Xie et al., 2004; Pan et al., 2006; Xie et al., 2010). The increased nitrogen content, SLA, and photosynthetic rate suggest that E. crassipes has a higher nutrition capture efficiency, resource use efficiency, and growth rate in high nutrient environments, which contribute to its superior performance compared with co-occurring native plants. Furthermore, although some studies found that invasive plants outperformed native plants in growth rate, P_{max} , resource use efficiency, and tissue nutrient content, in all habitats (Pattison et al., 1998; Durand & Goldstein, 2001), our study suggests that E. crassipes did not have leaf trait advantages over the native plant M. vaginalis at all three nutrient levels. Only at high nutrient levels, did it achieve significantly higher P_{max} and PNUE than the native species M. vaginalis. At the low nutrient level, the native species had significantly higher values in all the studied leaf traits (except PNUE), consistent with previous studies (Daehler, 2003; Xie et al., 2010). Therefore, our results suggest that the success of E. crassipes is context-dependent—it outperformed co-occurring native plants only in high nutrient environments. Some studies showed that E. crassipes was competitively superior at a range of resource availabilities. For instance, Xie (2003) found E. crassipes had a greater biomass and a higher phosphate absorbption rate than native aquatic plant Hydrocharis dubia (Bl.) Backer at all three phosphate levels. These different results may reflect the unique characteristics in the species or parameters in the studies. In fact, consistent with our results, E. crassipes was rare in Liangzi Lake, a nutrient poor lake (N and P concentrations in water were 0.71 and 0.04 mg l⁻¹; N concentration in sediment was 2.94 and 0.12 mg g⁻¹), and was only present in areas where wastewater was expelled.

Conclusion

After comparing multiple resource capture- and userelated traits of *E. crassipes* with *M. vaginalis* at three nutrient levels, we found the two plants have distinct morphological and physiological response patterns to changes in nutrients. The differences may be due to different strategies of reproduction and help explain the invasiveness of *E. crassipes*. Furthermore, our results supported the fluctuating resources hypothesis,



and found that increasing the nutrient content in sediment will augment the nutrition capture efficiency, resource use efficiency and growth rate of *E. crassipes*. Our results suggest a characteristic of content-dependency of *E. crassipes* invasion and explain the distribution patterns of *E. crassipes* in invaded areas: *E. crassipes* is prevalent in tropical and sub-tropical water bodies in which the water nutrient concentrations are often high due to agricultural runoff, deforestation, and insufficient wastewater treatment (Villamagna & Murphy, 2010).

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References

- Abbsi, S. A. & P. C. Nipaney, 1986. Infestation by aquatic weeds of the fern genus *Salvinia*: its status and control. Environmental Conservation 13: 235–241.
- Baruch, Z. & G. Goldstein, 1999. Leaf construction cost, nutrient concentration, and net CO₂ assimilation of native and invasive species in Hawaii. Oecologia 121: 183–192.
- Bloom, A. J., F. S. Chapin III & H. A. Mooney, 1985. Resource limitation in plants—an economic analogy. Annual Review of Ecology and Systematics 16: 363–392.
- Burns, J. H., 2004. A comparison of invasive and non-invasive dayflowers (*Commelinaceae*) across experimental nutrient and water gradients. Diversity and Distributions 10: 387– 397
- Chapin III, F. S., 1980. The mineral nutrition of wild plants. Annual Review of Ecology and Systematics 11: 233–260.
- Cohen, A. N. & J. T. Carlton, 1998. Accelerating invasion rate in a highly Invaded estuary. Science 279: 555–558.
- D'Antonio, C. M. & S. Kark, 2002. Impacts and extent of biotic invasions in terrestrial ecosystems. Trends in Ecology & Evolution 17: 202–204.
- Daehler, C. C., 2003. Performance comparisons of co-occurring native and alien invasive plants: implications for conservation and restoration. Annual Review of Ecology, Evolution and Systematics 34: 183–211.
- Davis, M. A., J. P. Grime & K. Thompson, 2000. Fluctuating resources in plant communities: a general theory of invasibility. Journal of Ecology 88: 528–534.
- Dukes, J. S. & H. A. Mooney, 1999. Does global change increase the success of biological invaders? Trends in Ecology & Evolution 14: 135–139.
- Durand, Z. L. & G. Goldstein, 2001. Photosynthesis, photoinhibition, and nitrogen use efficiency in native and invasive tree ferns in Hawaii. Oecologia 126: 345–354.
- Evans, J. R., 1989. Photosynthesis and nitrogen relationships in leaves of C3 plants. Oecologia 78: 9–19.

- Feng, Y. L., 2008. Photosynthesis, nitrogen allocation and specific leaf area in invasive *Eupatorium adenophorum* and native *Eupatorium japonicum* grown at different irradiances. Physiologia Plantarum 133: 318–326.
- Feng, Y. L. & G. L. Fu, 2008. Nitrogen allocation, partitioning and use efficiency in three invasive plant species in comparison with their native congeners. Biological Invasions 10: 891–902.
- Feng, Y. L., H. Auge & S. K. Ebeling, 2007. Invasive *Buddleja davidii* allocates more nitrogen to its photosynthetic machinery than five native woody species. Oecologia 153: 501–510.
- Feng, Y. L., G. L. Fu & Y. L. Zheng, 2008. Specific leaf area relates to the differences in leaf construction cost, photosynthesis, nitrogen allocation, and use efficiencies between invasive and noninvasive alien congeners. Planta 228: 383–390.
- Feng, Y. L., Y. B. Lei, R. F. Wang, R. M. Callaway, A. Valiente-Banuet, Y. P. Li Inderjit & Y. L. Zheng, 2009. Evolutionary tradeoffs for nitrogen allocation to photosynthesis versus cell walls in an invasive plant. Proceedings of the National Academy of Sciences 106: 1853–1856.
- Funk, J. L. & P. M. Vitousek, 2007. Resource-use efficiency and plant invasion in low-resource systems. Nature 446: 1079–1081.
- Garcia-Serrano, H., J. Escarre, E. Garnier & F. X. Sans, 2005. A comparative growth analysis between alien invader and native Senecio species with distinct distribution ranges. Ecoscience 12: 35–43.
- Grotkopp, E. & M. Rejmánek, 2007. High seedling relative growth rate and specific leaf area are traits of invasive species: phylogenetically independent contrasts of woody angiosperms. American Journal of Botany 94: 526–532.
- Grotkopp, E., M. Rejmánek & T. L. Rost, 2002. Toward a causal explanation of plant invasiveness: seedling growth and life history strategies of 29 pine (Pinus) species. The American Naturalist 159: 396–419.
- Howard, G. W. & K. L. S. Harley, 1998. How do floating aquatic weeds affect wetland conservation and development? How can these effects be minimised? Wetlands Ecology and Management 5: 215–225.
- Hulme, P. E., 2009. Trade, transport and trouble: managing invasive species pathways in an era of globalization. Journal of Applied Ecology 46: 10–18.
- Juliano, S. A. & L. P. Lounibos, 2005. Ecology of invasive mosquitoes: effects on resident species and on human health. Ecological Letters 8: 558–574.
- Leishman, M. R., T. Haslehurst, A. Ares & Z. Baruch, 2007. Leaf trait relationships of native and invasive plants: community- and global-scale comparisons. New Phytologist 176: 635–643.
- Lowe S., M. Browne, S. Boudjelas, & M. De Poorter (2000). 100 of the World's Worst Invasive Alien. Species A Selection from the Global Invasive Species Database. Invasive Species Specialist Group (ISSG), a specialist group of the Species Survival Commission (SSC) of the World Conservation Union (IUCN): 12 pp.
- Matzek, V., 2011. Superior performance and nutrient-use efficiency of invasive plants over non-invasive congeners in a resource-limited environment. Biological Invasions 13: 3005–3014.



- McAllister, C. A., A. K. Knapp & L. A. Maragni, 1998. Is leaf-level photosynthesis related to plant success in a highly productive grassland? Oecologia 117: 40–46.
- McDowell, S. C. L., 2002. Photosynthetic characteristics of invasive and noninvasive species of *Rubus* (Rosaceae). American Journal of Botany 89: 1431–1438.
- Niinemets, Ü., F. Valladares & R. Ceulemans, 2003. Leaf-level phenotypic variability and plasticity of invasive *Rhodo-dendron ponticum* and non-invasive *Ilex aquifolium* co-occurring at two contrasting European sites. Plant, Cell & Environment 26: 941–956.
- Njambuya, J. & L. Triest, 2010. Comparative performance of invasive alien *Eichhornia crassipes* and native *Ludwigia* stolonifera under non-limiting nutrient conditions in Lake Naivasha, Kenya. Hydrobiologia 656: 221–231.
- Ogutu-Ohwayo, R., R. E. Hecky, A. S. Cohen & L. Kaufman, 1997. Human impacts on the African Great Lakes. Environmental Biology of Fishes 50: 117–131.
- Osunkoya, O. O., D. Bayliss, F. D. Panetta & G. Vivian-Smith, 2010. Leaf trait co-ordination in relation to construction cost, carbon gain and resource-use efficiency in exotic invasive and native woody vine species. Annals of Botany 106: 371–380.
- Pan, X., Y. Geng, W. Zhang, B. Li & J. Chen, 2006. The influence of abiotic stress and phenotypic plasticity on the distribution of invasive *Alternanthera philoxeroides* along a riparian zone. Acta Oecologica 30: 333–341.
- Parker, J. D. & M. E. Hay, 2005. Biotic resistance to plant invasions? Native herbivores prefer non-native plants. Ecology Letters 8: 959–996.
- Pattison, R. R., G. Goldstein & A. Ares, 1998. Growth, biomass allocation and photosynthesis of invasive and native Hawaiian rainforest species. Oecologia 117: 449–459.
- Pimentel, D., R. Zuniga & D. Morrison, 2005. Update on the environmental and economic costs associated with alieninvasive species in the United States. Ecological Economics 52: 273–288.
- Poorter, H. & J. R. Evans, 1998. Photosynthetic nitrogen-use efficiency of species that differ inherently in specific leaf area. Oecologia 116: 26–37.

- Shipley, B., 2006. Net assimilation rate, specific leaf area and leaf mass ratio: which is most closely correlated with relative growth rate? A eta-analysis. Functional Ecology 20: 565–574.
- Svilà, M., J. L. Espinar, M. Hejda, P. E. Hulme, V. Jarošík, J. L. Maron, J. Pergl, U. Schaffner, Y. Sun & P. Pyšek, 2011. Ecological impacts of invasive alien plants: a metaanalysis of their effects on species, communities and ecosystems. Ecological Letters 14: 702–708.
- Villamagna, A. M. & B. R. Murphy, 2010. Ecological and socioeconomic impacts of invasive water hyacinth (*Eichhornia* crassipes): a review. Freshwater Biology 55: 282–298.
- Vitousek, P. M., 1986. Biological invasions and ecosystem properties: can species make a difference? Ecological Studies 58: 163–176.
- Vitousek, P. M., 1990. Biological invasions and ecosystem processes: towards an integration of population biology and ecosystem studies. Oikos 57: 7–13.
- Xie, Y. H. (2003). Studies on the Nutrient Ecology of Water Hyacinth (*Eichhornia crassipes*), an Exotic Invasive Species. Ph.D. thesis, Wuhan University (in Chinese with English abstract).
- Xie, Y. H. & D. Yu, 2003. The significance of lateral roots in phosphorus (P) acquisition of water hyacinth (*Eichhornia* crassipes). Aquatic Botany 75: 311–321.
- Xie, Y. H., M. Z. Wen, D. Yu & Y. k. Li, 2004. Growth and resource allocation of water hyacinth as affected by gradually increasing nutrient concentrations. Aquatic Botany 79: 257–266.
- Xie, D., D. Yu, L. F. Yu & C. H. Liu, 2010. Asexual propagations of introduced exotic macrophytes *Elodea nuttallii*, *Myriophyllum aquaticum*, and *M. propinquum* are improved by nutrient-rich sediments in China. Hydrobiologia 655: 37–47.
- Xiong, W., D. Yu, Q. Wang, C. H. Liu & L. G. Wang, 2008. A snail prefers native over exotic freshwater plants: implications for the enemy release hypotheses. Freshwater Biology 53: 2256–2263.

